

Aqueous Extract Of Tutsan (*Hypericum Androsaemum L.*) Red Berries Exhibits Antioxidant Activity And Effects Akin To Those Of An Antidepressant

Satish Kilaje^{1*}, Laxmikant Purane², Sonal Barde³, Varsha Patwekar⁴, Shyam Awate⁵, Shailesh Patwekar⁶, Santosh Butle⁷

^{1*}Dr. J. J. Magdum Pharmacy College, Jaysingpur Tal. Shirol, Dist. Kolhapur

²Department of Pharmacology, YSPM's, Yashoda Technical Campus, Faculty of Pharmacy, Satara

³Assistant Professor, Sinhagad Institute of Pharmacy, Vadgaon, Pune

⁴Dr Vedprakash Patil college of pharmacy, Aurangabad.

⁵Assistant Professor, Indrayani Institute of Pharmaceutical Education and Research, Talegaon Dabh., Pune.

⁶Assistant Professor, School of Pharmacy, SRTM University, Nanded

⁷Associate Professor, School of Pharmacy, SRTM University, Nanded.

***Corresponding Author:** Satish Kilaje

***Dr. J. J. Magdum Pharmacy College, Jaysingpur Tal. Shirol, Dist. Kolhapur**

Abstract:

Hypericum androsaemum L., sometimes referred to as "tutsan" or "shrubby St. John's Wort," is a plant in the *Hypericum* genus that grows naturally in the Mediterranean region. It is widely grown as an ornamental plant because of the striking color variation in its young, berry-like capsules, which turn shiny black as they ripen from red. In traditional medicine from Portugal and Spain, tutsan has also been used to treat depression. We evaluated the potential benefits of *H. androsaemum* red berry water extract (WE). After *H. androsaemum* red berries were decocted, WE was produced, and its ten bioactive component content was ascertained using HPLC-DAD analysis. By boosting endogenous antioxidant defenses and restoring normal behavior, the maximum dosage of WE was found to effectively alleviate the symptoms of depression overall and lower oxidative stress levels.

Keywords: Tutsan, Red berries, Antidepressant, *Hypericum androsaemum L.*, etc.

Introduction:

Stroke is one of the most significant cerebrovascular illnesses, with high global death and disability rates ^[1]. There are two types of stroke: ischemic and hemorrhagic. The latter results in brain damage and infarction, and cerebral blood flow is compromised by low blood sugar and oxygen levels ^[2]. Reactive oxygen species (ROS), which lead to neuronal malfunction and death, are produced as a result, changing antioxidant defenses ^[3]. Major depressive disorder (MDD) is characterised by severe biological molecular damage that leads to neurotoxicity and neurodegeneration [5], and this oxidative stress plays a role in its development [4]. Therefore, common symptoms that follow an acute ischemic stroke include anxiety, depression, psychosis, apathy, dementia, and chronic fatigue syndrome. ^[6]. Depression is one of these, affecting stroke patients frequently and being a major neuropsychiatric condition ^[7]. Poor recovery outcomes, social issues, isolation, cognitive impairment, sleeplessness, and an increased risk of death are common symptoms of post-stroke depression (PSD) ^[6, 8, 9].

The primary therapeutic approach for treating PSD is the use of antidepressant medications; however, these frequently have substantial side effects that can aggravate morbidity and even cause therapy to be stopped ^[10]. PSD clinical symptoms bear resemblance to those observed in major depressive disorder cases. Thus, oxidative stress is crucial for both major depressive disorder and associated psychiatric diseases, as well as chronic inflammation ^[11, 12]. We have previously postulated that oxidative stress is a significant factor in stroke-induced damage as well as serious depressive disorders ^[6, 13]. Our search for herbal remedies with antioxidant and antidepressant qualities that could be clinically useful in the treatment of depression following a stroke led us to concentrate on the *Hypericum* genus (*Hypericaceae* family). A variety of conventional herbal medications and dietary supplements contain extracts from the aerial portions of *Hypericum perforatum L.*, which are commonly used as a depression treatment ^[14]. Known as "tutsan" or "shrubby St. John's Wort," *Hypericum androsaemum* (sect. *Androsaemum*) has been little research done on its pharmacological properties to date. The Mediterranean shrub Tutsan grows in forests where oak and chestnut trees predominate. *H. androsaemum* is referred to as "iperico arbustivo," "ruta sana," "erba di San Giovanni arbustiva," and other local terms in Italy ^[15]. *H. androsaemum* has fleshy, berry-like capsules that turn black as they ripen, in contrast to most other *Hypericum* species, which have dry capsules as fruit ^[16, 17]. Another characteristic that sets Tutsan plants apart is their lack of black leaf nodules, which are secretory structures where naphthodianthrones are formed [18]. Consequently,

hypericin and its compounds are absent from the resultant plant extracts [16]. In the Mediterranean region, *Hypericum androsaemum* is one of the most widely used herbal remedies in the genus, behind St. John's Wort. While its aerial sections are infused and used to treat anxiety and depression in Spain [22], it is known as "Hipericao do Geres" in Portugal and is used as a hepatoprotective, diuretic, and antidepressant [19–21]. While methanolic extracts from *H. androsaemum* aerial parts have been shown to have depressive qualities in vivo [21], the fruits' potential for medicinal application has not been investigated. In a previous study, Based on our earlier research, we postulated that the antioxidant capacity of berries like *H. androsaemum* could be beneficial in managing post-stroke depression symptoms. Thus, we tested water extracts from *H. androsaemum* red berries for possible antidepressant and antioxidant benefits in post-stroke depressive-like effects in mice brains.

Material and Method Plant material

In July, red berries were harvested from *Hypericum androsaemum* L. This stage of the fruit provides the maximum concentrations of bioactive components, such as phenolic acids and flavonoids, according to earlier studies [16, 23].



Fig. 1. Rosemary with hypericum spp. berry-like red capsules that were examined in the research Reagents and standards

The following analytical reference samples were acquired from Sigma-Aldrich: hyperoside, quercitrin, rutin, 3, 5-di-O-caffeoylquinic acid, chlorogenic acid, neochlorogenic acid, shikimic acid, epicatechin, and gallic acid. Vials were stored at 4 °C in the dark after each analyte was dissolved in methanol to create stock standard solutions with a concentration of 1000 mg/1000 mL. Every day, new standard solutions were made. Sigma-Aldrich supplied HPLC-grade acetonitrile and methanol, while J.T. Baker B.V. supplied HPLC-grade formic acid. Deionized water ≥ 18 M Ω /cm resistivity, purified using a Milli-Q system was utilized for the preparation of samples and chromatographic analyses. Furthermore, Supelco supplied 0.45- μ m PTFE filters, which were used to filter all solvents and solutions prior to usage.

Preparations of water extracts

After macerating 50 g of crushed red fruits in 300 mL of boiling water for 30 minutes, water extract (WE) was produced (producing 17.09 %, w/w dry weight). Using a Rota vapor, the extract was concentrated, lyophilized, and stored at 4 °C in the freezer until needed. Prior to chromatographic analysis, 10 mg of extract were filtered out of a sample solution made up of 1 mL of water for HPLC analysis.

Biochemical investigation Lipid peroxidation

Thiobarbituric acid-reactive substance (TBARS), a marker for lipid peroxidation, was quantified using a method developed by our group [29, 30]. Samples of homogenised brain tissue with 1 mg of protein were mixed with about 1 mL of 20% trichloroacetic acid and 2 mL of 0.67% thiobarbituric acid. After an hour of incubation at 100 °C, the mixture was cooled and the precipitate was removed by centrifuging the samples. The absorbance of the reagents, excluding brain homogenate, was measured at $\lambda = 532$ nm in reference to a blank in order to provide a control for the reaction mixtures.

Superoxide dismutase

We used the Misra and Fridovich [31] procedure to measure the activity of superoxide dismutase (SOD). In short, 0.2 mL of freshly prepared hydroxylamine hydrochloride (0.1 mM), 0.4 mL of nitroblue tetrazolium (25 M), and 1 mL of sodium carbonate (50 mM) were combined to create a combination. Next, 0.1 mL of the clear supernatant of the homogenate (1:10, % w/v) was added. Variations in absorbance were measured at $\lambda = 560$ nm.

Reduced glutathione levels

Glutathione (GSH) level drop was examined using Ellman's approach [32]. The protein was precipitated using trichloroacetic acid (5%) after homogenates (720 μ L) were diluted twice. The supernatant was collected following centrifugation (12,000 g, 5 min), and Ellman's reagent (5, 5-dithiobis 2-nitrobenzoic acid solution) was then added. The absorbance of each sample was then determined at $\lambda = 417$ nm. Next, a standard curve was created using reduced glutathione solution concentrations that were known.

Catalase activity

In this work, a slightly modified version of Sinha's [33][29] technique was used to quantify catalase activity. In summary, 2.1 mL of 7.5 mM hydrogen peroxide was combined with a homogenate volume containing 5 μ g of protein. The absorbance of the reaction mixture was measured at $\lambda = 240$ nm after 10 minutes.

Statistical analysis

IBM SPSS 21.0 for Windows, a statistical application for the social sciences, was used for statistical analysis. The results were shown as means \pm standard deviation (SD), and statistical significance was established at $p < 0.05$ using a one-way variance analysis. The Bonferroni post-hoc test was used to find any significant differences between the groups.

Result and Discussion

HPLC analysis of the water extract

For this study, ten bioactive marker molecules were evaluated in the red berry *H. androsaemum* water extract (WE) (Table 1). The target analytes represented 180.6 g/kg of WE, of which the three main components were catechin (5.8 g/kg), chlorogenic acid (56.9 g/kg), and shikimic acid (110.0 g/kg) (Fig. 2). Hyperoside, another common secondary metabolite, was discovered in flavonol glycoside at a concentration of 2.7 g/kg.

Table 1: Marker component content in the *Hypericum androsaemum* red berry water extract

Compound	Content (g/kg dw)
Shikimic acid	111.1 \pm 3.0
Gallic acid	0.6 \pm 0.03
Neochlorogenic acid	1.1 \pm 0.02
Chlorogenic acid	57.8 \pm 1.3
3,5-di-O-caffeoylquinic acid	n.d.
Catechin	5.9 \pm 0.4
Epicatechin	2.1 \pm 0.09
Rutin	1.7 \pm 0.2
Hyperoside	2.8 \pm 0.09
Quercitrin	n.d.
Total	181.7 \pm 1.6

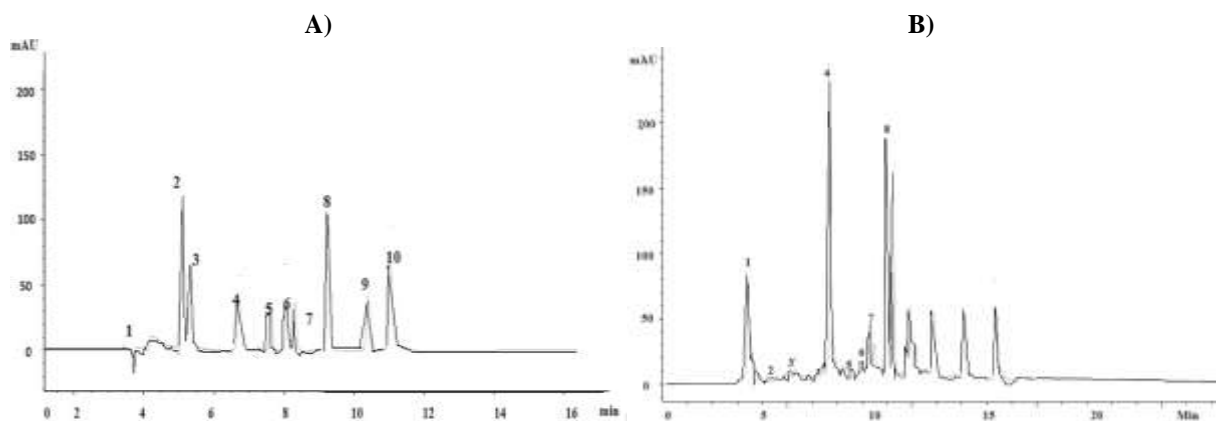


Fig. 2: Chromatographs using HPLC-DAD. They are given as follows for the purpose of clarity: A) reference mixture solution containing the ten analytes under survey, B) aqueous extract of *H. androsaemum*. The measurements are taken at 256 nm. List of substances 1, 2, gallic acid, 3, 5-O-caffeoylquinic acid, 4 3-O-caffeoylquinic acid, 5 epicatechin, 6 catechin, 7 rutin, 8 hyperoside, 9 quercitrin, and 10 3,5-di-O-caffeoylquinic acid.

After administering WE at both tested doses, the antioxidant properties of WE were assessed. The induction of stroke led to an increase in TBARS levels and a significant ($p < 0.01$) decrease in the activity of endogenous antioxidant defenses, including SOD, GSH, and Cat. (Fig. 6A–D). The extracts exhibited impressive antioxidant activity, despite their inability to return these endogenous antioxidant molecules to normal physiological quantities. At the higher concentration tested (30 mg/kg), WE was determined to be the most effective treatment (Fig. 6A–D).

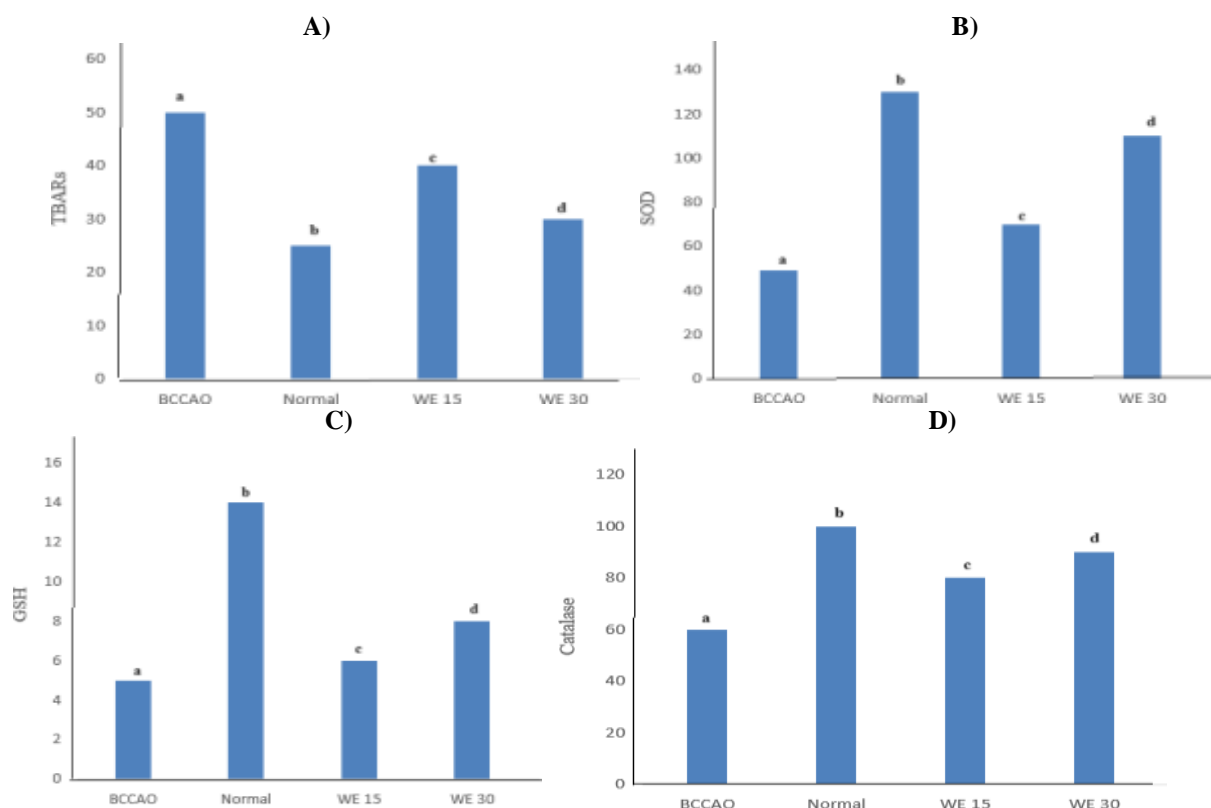


Fig. The impact of red berry extracts (WE) from *Hypericum* and *Rosaemum* on oxidative stress. Intraperitoneal WE administration's effects on the following parameters in mouse brain tissue: glutathione (GSH) levels, expressed as $\mu\text{g}/\text{mg}$ protein (C), catalase activity, expressed as U/mg protein (D), superoxide dismutase (SOD) activity, expressed as nmol MDA equivalent/g tissues (A), and Glutathione (TBARS) levels, expressed as $\mu\text{g}/\text{mg}$ protein (B). The data are presented as means \pm SD ($n = 7$); significant differences ($p < 0.05$) between the two groups are indicated by different letters.

The relationships between the antioxidant activity and the antidepressant-like effects found in behavioral data were investigated using Pearson's linear correlation. Antioxidant enzymes actually demonstrated a high positive link with both movement metrics, but TBARS exhibited a substantial negative relationship (Table 2).

Conclusion

The polar extract from the fruit of *H. androsaemum*, a well-known Mediterranean medicinal plant, has been shown in this work to have health-promoting properties. We expect that more research will uncover this portion of the *H. androsaemum*'s potential, which may subsequently be used to make nutraceuticals and functional foods that protect against oxidative stress and neurodegenerative illnesses.

References

1. V. Vukovic, I. Mikula, M.J. Kestic, M.R. Bedekovic, S. Morovic, A. Lovrencic-Huzjan, V. Demarin. Perception of stroke in Croatia—knowledge of stroke signs and risk factors amongst neurological outpatients. *Eur. J. Neurol.* 2009; 16: 1060–1065.
2. D. Lloyd-Jones, R. Adams, M. Carnethon, G. De Simone, T.B. Ferguson, K. Flegal, E. Ford, K. Furie, A. Go, K. Greenlund. Heart disease and stroke statistics—2009 update a report from the American Heart Association Statistics Committee and Stroke Statistics Subcommittee. *Circulation.* 2009; 119; 21–181.
3. K. Niizuma, H. Endo, P.H. Chan. Oxidative stress and mitochondrial dysfunction as determinants of ischemic neuronal death and survival. *J. Neurochem.* 2009; 109; 133–138.
4. E.H. Tobe. Mitochondrial dysfunction, oxidative stress, and major depressive disorder, *Neuropsychiatr. Dis. Treat.*

- 2013;9: 567-557.
5. F. Ng, M. Berk, O. Dean, A.I. Bush. Oxidative stress in psychiatric disorders: evidence base and therapeutic implications, *Int. J. Neuropsychopharmacol.* 2008; 11: 851–876.
 6. S.-F. Nabavi, O.-M. Dean, A. Turner, A. Sureda, M. Daglia, S.-M. Nabavi. Oxidative stress and post-stroke depression: possible therapeutic role of polyphenols? *Curr. Med. Chem.* 2015; 22: 343–351.
 7. M.L. Hackett, K. Pickles. Part I: frequency of depression after stroke: an updated systematic review and meta-analysis of observational studies. *Int. J. Stroke.* 2014; 9: 1017–1025.
 8. S.-F. Nabavi, S. Habtemariam, A. Di Lorenzo, A. Sureda, S. Khanjani, S.-M. Nabavi, M. Daglia. Post-stroke depression modulation and in vivo antioxidant activity of gallic acid and its synthetic derivatives in a murine model system. *Nutrients.* 2016; 8: 248.
 9. N. Hadidi, D.J. Treat-Jacobson, R. Lindquist. Post stroke depression and functional outcome: a critical review of literature. *Heart Lung.* 2009; 38:151–162.
 10. I. Loubinoux, G. Kronenberg, M. Endres, P. Schumann-Bard, T. Freret, R.K. Filipkowski, L. Kaczmarek, A. Popa-Wagner. Post-stroke depression: mechanisms, translation and therapy. *J. Cell. Mol. Med.* 2012; 16: 1961–1969.
 11. J.D. Rosenblat, D.S. Cha, R.B. Mansur, and R.S. McIntyre. Inflamed moods: a review of the interactions between inflammation and mood disorders, *Prog. Neuro Psychopharmacol. Biol. Psychiatry.* 2014; 53: 23–34.
 12. J.D. Rosenblat, E. Brietzke, R.B. Mansur, N.A. Maruschak, Y. Lee, R.S. McIntyre. Inflammation as a neurobiological substrate of cognitive impairment in bipolar disorder: evidence, pathophysiology and treatment implications. *J. Affect. Disord.* 2015; 188; 149–159.
 13. A. Di Lorenzo, S.F. Nabavi, A. Sureda, A.H. Moghaddam, S. Khanjani, P. Arcidiaco, S.M. Nabavi, M. Daglia. Antidepressive-like effects and antioxidant activity of green tea and GABA green tea in a mouse model of post-stroke depression. *Mol. Nutr. Food Res.* 2016; 60: 566–579.
 14. E.A. Apaydin, A.R. Maher, R. Shanman, M.S. Booth, J.N. Miles, M.E. Sorbero, S. Hempel. A systematic review of St. John's Wort for major depressive disorder. *Syst. Rev.* 2016; 5: 148.
 15. S. Ballelli, B. Bellomaria, *La Flora officinale delle Marche, l'Uomo e l'ambiente-43*, Camerino, Arte Lito vol. 1, (2005), pp. 174–176.
 16. G. Caprioli, A. Alunno, D. Beghelli, A. Bianco, M. Bramucci, C. Frezza, R. Iannarelli, F. Papa, L. Quassinti, G. Sagratini, B. Tirillini, A. Venditti, S. Vittori, F. Maggi. Polar constituents and biological activity of the berry-like fruits of *Hypericum androsaemum*. *Front. Plant Sci.* 2016; 7: 232.
 17. G. Caprioli, R. Iannarelli, K. Cianfaglione, D. Fiorini, C. Giulliani, D. Lucarini, F. Papa, G. Sagratini, S. Vittori, F. Maggi. Volatile profile, nutritional value and secretory structures of the berry-like fruits of *Hypericum androsaemum* L. *Food Res. Int.* 2016; 79: 1–10.
 18. S.L. Crockett, N.K.B. Robson. Taxonomy and chemotaxonomy of the genus *Hypericum*. *Med. Aromat. Plant. Sci. Biotechnol.* 2011; 5: 1–13.
 19. A.M. Carvalho, *Plantas y sabiduria popular del Parqueparque Natural de Montesinho, Un estudio etnobotánico en Portugal*, Consejo Superior De Investigaciones Cientificas (CSIC), Madrid, Spain, 2010.
 20. P. Valentao, E. Fernandes, F. Carvalho, P.B. Andrade, R.M. Seabra, M.D.L. Bastos. Antioxidant activity of *Hypericum androsaemum* infusion scavenging activity against superoxide radical, hydroxyl radical and hypochlorous acid. *Biol. Pharm. Bull.* 2002; 25: 1320–1323.
 21. N. Ramalhete, A. Machado, R. Serrano, E.T. Gomes, H. Mota-Filipe, O. Silva. Comparative study on the in vivo antidepressant activities of the Portuguese *Hypericum foliosum*, *Hypericum androsaemum* and *Hypericum perforatum* medicinal plants. *Ind. Crops Prod.* 2016; 82: 29–36.
 22. S. Akerreta, M.I. Calvo, R.Y. Caverro, *Sabiduría popular y plantas curativas*, Ediciones I, Integralia La casa natural S.L., Madrid, Spain, 2013.
 23. F. Antognoni, M. Lianza, F. Poli, M. Buccioni, C. Santinelli, G. Caprioli, R. Iannarelli, G. Lupidi, E. Damiani, D. Beghelli, A. Alunno, F. Maggi. Polar extracts from the berry-like fruits of *Hypericum androsaemum* L. as a promising ingredient in skin care formulations, *J. Ethnopharmacol.* 2017; 195; 255–265.
 24. V. Lopez, F. Res, R. Iannarelli, G. Caprioli, F. Maggi. Methanolic extract from red berry-like fruits of *Hypericum androsaemum*: chemical characterization and inhibitory potential of central nervous system enzymes. *Ind. Crops Prod.* 2016; 94: 363–367.
 25. S.-F. Nabavi, E. Sobarzo-Sanchez, S.-M. Nabavi, M. Daglia, A.H. Moghaddam, A.G. Silva. Behavioral effects of 2, 3- dihydro-and oxoisoaporphine derivatives in post stroke-depressive like behavior in male balb/c mice. *Curr. Topics Med. Chem.* 2013; 13: 2127–2133.
 26. A. Hajizadeh Moghaddam, E. Sobarzo-Sanchez, S.-F. Nabavi, M. Daglia, S. - M. Nabavi. Evaluation of the antipsychotic effects of 2-(dimethylamino)-and 2- (methylamino)-7H-naphtho [1, 2, 3-de] quinolin-7-one derivatives in experimental model of psychosis in mice. *Curr. Topics Med. Chem.* 2014; 14: 229–233.
 27. S.-F. Nabavi, E. Sobarzo-Sanchez, S.-M. Nabavi, A. Sureda, A. Hajizadeh Moghaddam. Bi-3-aza-oxoisoaporphine derivatives have antidepressive properties in a murine model of post stroke-depressive like behavior. *Curr.*

- Neurovasc. Res.2013; 10: 164–171.
28. M.-M. Bradford. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal. Biochem.* 1976; 72: 248–254.
 29. S.-M. Nabavi, S.-F. Nabavi, S. Eslami, A.-H. Moghaddam. In vivo protective effects of quercetin against sodium fluoride-induced oxidative stress in the hepatic tissue. *Food Chem.* 2012; 132: 931–935.
 30. S.-F. Nabavi, S.-M. Nabavi, M. Mirzaei, A.-H. Moghaddam. Protective effect of quercetin against sodium fluoride induced oxidative stress in rat's heart. *Food Funct.* 2012; 3: 437–441.
 31. H.-P. Misra, I. Fridovich. The role of superoxide anion in the autoxidation of epinephrine and a simple assay for superoxide dismutase. *J. Biol. Chem.* 1972; 247: 3170–3175.
 32. G.-L. Ellman. Tissue sulfhydryl groups. *Arch. Biochem. Biophys.* 1959; 82: 70–77.
 33. A.K. Sinha. Colorimetric assay of catalase. *Anal. Biochem.* 1972; 47: 389–394.
 34. SL Patwekar, MK Baramade. Controlled release approach to novel multiparticulate drug delivery system. *Int J Pharm Pharm Sci* 4 (3), 757-63
 35. SL Patwekar, AB Suryawanshi, MS Gaikwad, SR Pedewad, AP Potulwar. Standardization of herbal drugs: An overview. *The Pharma Innovation* 5 (4, Part B), 100.
 36. SL Patwekar, SG Gattani, MM Pande. Needle free injection system: A review. *Int J Pharm Pharm Sci* 5 (4), 14-19.
 37. JDS Khayyam Shaikh , Shailesh Patwekar , Santosh Payghan. Dissolution and Stability Enhancement of Poorly Water Soluble Drug – Lovastatin by Preparing Solid Dispersions. *Asian Journal of Biomedical and Pharmaceutical Sciences* 1 (4), 24-31
 38. PG Jamkhande, VA Suryawanshi, TM Kaylankar, SL Patwekar. Biological activities of leaves of ethnomedicinal plant, *Borassus flabellifer* Linn.(Palmyra palm): An antibacterial, antifungal and antioxidant evaluation. *Bulletin of Faculty of Pharmacy, Cairo University* 54 (1), 59-66.
 39. SL Patwekar, SR Pedewad, S Gattani. Development and evaluation of nanostructured lipid carriers-based gel of isotretinoin. *Particulate Science and Technology* 36 (7), 832-843.
 40. PP Sambarkar, SL Patwekar, BM Dudhgaonkar. Polymer nanocomposites: An overview. *Int J Pharm Pharm Sci* 4 (2), 60-65.
 41. SL Patwekar. Nanobiocomposite: A new approach to drug delivery system. *Asian Journal of Pharmaceutics (AJP)* 10 (04).
 42. PM Dhere, SL Patwekar. Review on preparation and evaluation of oral disintegrating films. *Int J Pharm Tech* 3 (4), 1572-1585.
 43. S Patwekar, S Gattani, R Giri, A Bade, B Sangewar, V Raut. Review on nanoparticles used in cosmetics and dermal products. *World J. Pharm. Pharm. Sci* 3, 1407-1421.
 44. SA Payghan, VK Kate, K Khavane, SS Purohit. Pharmaceutical solid polymorphism: Approach in regulatory consideration. *J Glob Pharm Technol* 1, 45-53.
 45. L Shailesh, RP Snehal, P Ashwini, S Manoj, B Arvind. Nanostructured lipid carriers in stability improvement for cosmetic nanoparticles. *International Journal of Pharmacy & Pharmaceutical Research* 6 (1), 168-180.
 46. SL Patwekar, RS Sakhare, NN Nalbalwar. HPLC method development and validation-A general Concept. *International Journal of Chemical and Pharmaceutical Sciences* 6 (1), 8-14.
 47. SPD V.N Gunjkar, S.L.Patwekar. Stimuli Responsive Layer By Layer Self-Assembly A Novel Approachs In Current Drug Delivery: Review. *World Journal of Pharmacy And Pharmaceutical Sciences* 4 (6), 216-238.
 48. SL Patwekar. Solubility and dissolution enhancement of poorly water-soluble Ketoprofen by microwave-assisted Bionanocomposites: in vitro and in vivo study. *Asian Journal of Pharmaceutics (AJP)* 10 (04).
 49. KA Nangare, SD Powar, VK Kate, SR Patwekar, SA Payghan. Therapeutics Applications of Nanosuspension in Topical/Mucosal Drug Delivery. *Journal of Nanomedicine Research* 7 (1).
 50. K Khavane, V Addepalli, K Bhusare, SA Payghan, S Patwekar, V Kate. Prescribing patterns of antibiotics and sensitivity patterns of micro-organisms towards different antibiotics in multidisciplinary health care hospital. *International Journal of Pharmaceutical and Biologic Archives* 1 (2), 115-22.
 51. SG Gattani, SL Patwekar. Enhancement solubility and dissolution Rate of Ibuprofen by Nanobiocomposites using Microwave Induced Diffusion (MIND) Method. *World Journal Of Pharmacy and Pharmaceutical Sciences* 6 (11), 716-740.
 52. A Jirage, K Shaikh, K Vaishali, SA Payghan, S Patwekar. In vitro-in vivo correlation for poly (3-hydroxybutyrate) base ibuprofen extended release tablets. *Asian J. Pharm* 11, 18-26.
 53. S Patwekar, G Gattani, R Sakhare, A Khan, S Gaikwad, S Pedewad. Current features of USFDA and EMA process validation guidance. *Int. J. Pharm. Pharm. Res* 6 (1), 300-313.
 54. PG Jamkhande, SR Barde, SL Patwekar, PS Tidke. Plant profile, phytochemistry and pharmacology of *Cordia dichotoma*, 1009-16.
 55. GV Gole, SL Patwekar, A Doiphode, A Rode, S Shaikh. A Overview on Nanosponges. *A & V Publications* 12 (3), 210-212.

56. L Mahajan, N Kapase Sachin, G Sonawane, S Barde, R Sakhare, R Moon. The Highlights On Herbs Acts As An Anti-Cancer Property—A Systematic Review. *Natural Volatiles & Essential Oils Journal*, 15692-15704.
57. VK Magar, L Sonawane, S Patwekar. Molecular Docking Study Of Few Novels Pyrimidine Derivatives On Validated Target Enoyl Acyl Coa Reductase. *Latin American Journal of Pharmacy: A Life Science Journal* 42 (3), 777-791.
58. SL Patwekar, G Namdev, V Gole, A Rode, S Shaikh. A Overview on Nanoemulsion. *Asian Journal of Research in Pharmaceutical Sciences* 12 (3), 239-244.
59. AR Doiphode, SL Patwekar, N Guhade, V Gole, A Rode, S Shaikh. A Overview on nanoemulsion. *A & V Publications* 12 (3), 239-244.
60. MRP Rao, S Taktode, SS Shivpuje, S Jagtap. Optimization of Transmucosal Buccal Delivery of Losartan Potassium using Factorial Design. *Indian Journal of Pharmaceutical Education and Research*, 2016; 50(2): S132-S139.
61. N Patre, S Patwekar, S Dhage, S Shivpuje. Formulation & Evaluation Of Piroxicam Bionanocomposite For Enhancement of Bioavailability. *European Journal of Molecular & Clinical Medicine*, 2020; 7(11): 9362-9376.
62. SJ Wadher, SL Patwekar, SS Shivpuje, SS Khandre, SS Lamture. Stability Indicating Assay Methods for Simultaneous Estimation of Amoxicillin Trihydrate And Cloxacillin Sodium in Combined Capsule Dosage Form by UV-Spectrophotometric Method. *European Journal of Biomedical and Pharmaceutical sciences*, 2017; 4(10): 858-864.
63. Santosh A. Payghan Shivraj S. Shivpuje Shailesh L. Patwekar, Karna B. Khavane, Padmavati R. Chainpure. A Review on Different Preparation Method Used For Development of Curcumin Nanoparticles. *International Journal of Creative Research Thoughts*, 2021;9(1):4088-4101.
64. Zeba Ashfaq Sheikh P. R. Chainpure, S. L. Patwekar, S. S. Shivpuje. Formulation and evaluation of Garciniacambogia and Commiphoramukul Herbal tablets used for AntiObesity. *International Journal of Engineering, Science and Mathematics*, 2019; 8(4): 180-195.
65. Pravin P Karle, Shashikant C Dhawale, Vijay V Navghare, Shivraj S Shivpuje. Optimization of extraction conditions and evaluation of Manilkara zapota (L.) P. Royen fruit peel extract for in vitro α -glucosidase enzyme inhibition and free radical scavenging potential. *Future Journal of Pharmaceutical Sciences*, 2021; 7(1):1-10.
66. Sheetal Rathod P. R. Chainpure, S. L. Patwekar, S. S. Shivpuje. A Study Of Carica Papaya Concerning It's Ancient And Traditional Uses - Recent Advances And Modern Applications For Improving The Milk Secretion In Lactating Womens. *International Journal of Research*, 2019;8(2):1851-1861.
67. Shivraj S. Shivpuje Shailesh J. Wadher, Bhagwan B. Supekar. Development And Validation Of New Ft-Ir Spectrophotometric Method For Simultaneous Estimation Of Ambroxol Hydrochloride And Cetirizine Hydrochloride In Combined Pharmaceutical. *International Research Journal of Pharmacy*, 2019; 10(3):110-114.
68. Shivraj S. Shivpuje, Shailesh J. Wadher, Bhagwan B. Supekar. Simultaneous Estimation of Ambroxol Hydrochloride and Cetirizine Hydrochloride in Combined Solid Tablet Formulations by HPTLC- Densitometric Method. *Asian Journal of Biochemical and Pharmaceutical Research*, 2019; 9(1):1-10.
69. JW Sailesh, SS Shivraj, SI Liyakat. Development and Validation of Stability Indicating RP-HPLC Method for the Estimation of Simvastatin in Bulk and Tablet Dosage form. *Research Journal of Pharmacy and Technology*, 2018; 11(4): 1553-1558.
70. Patil S. S. Shivpuje Shivraj S. Patre Narendra G. Development and Validation Of Stability Indicating HPTLC Method For Determination of Nisoldipine (Niso) In Tablet Dosage Form. *European Journal of Biomedical and Pharmaceutical sciences*, 2017; 4(12):462468.
71. W Shailesh, K Tukaram, S Shivraj, L Sima, K Supriya. Development and Validation of Stability Indicating UV Spectrophotometric Method for Simultaneous Estimation of Amoxicillin Trihydrate and Metronidazole In Bulk And In-House Tablet. *World Journal of Pharmaceutical and Medical Research*, 2017;3(8):312-318.
72. J Wadher Shailesh, M Kalyankar Tukaram, S Shivpuje Shivraj. Development and Validation of Stability Indicating Assay Method for Simultaneous Estimation of Amoxicillin Trihydrate and Cloxacillin Sodium In Pharmaceutical Dosage Form By Using RP-HPLC. *World Journal of Pharmaceutical Research*, 2017; 10(6):1002-1006.
73. Shital S. Sangale, Priyanka S. Kale, Rachana B. Lamkane, Ganga S. Gore, Priyanka B. Parekar, Shivraj S. Shivpuje (2023). Synthesis of Novel Isoxazole Derivatives as Analgesic Agents by Using Eddy's Hot Plate Method. *South Asian Res J Pharm Sci*, 5(1): 18-27.
74. Priyanka B. Parekar, Shivraj S. Shivpuje, Vijay V. Navghare, Manasi M. Savale, Vijaya B. Surwase, Priti S. Mane-Kolpe, Priyanak S. Kale. Polyherbal Gel Development And Evaluation For Antifungal Activity, *European Journal of Molecular & Clinical Medicine*. 2022; 9(03): 5409-5418.
75. Jain AA, Mane-Kolpe PD, Parekar PB, Todkari AV, Sul KT, Shivpuje SS. Brief review on Total Quality Management in Pharmaceutical Industries, *International Journal of Pharmaceutical Research and Applications*. 2022; 7(05):1030-1036.
76. Sumaiyya. K. Attar, Pooja P. Dhanawade, Sonali S. Gurav , Perna H. Sidwadkar , Priyanka B. Parekar, Shivraj S. Shivpuje. Development and Validation of UV Visible Spectrophotometric Method for Estimation of Fexofenadine Hydrochloride in Bulk and Formulation, *GIS SCIENCE JOURNAL*. 2022; 9(11): 936-944.

77. Sumayya Kasim Atar, Priyadarshini Ravindra Kamble, Sonali Sharad Gurav, Pooja Pandit Dhanawade, Priyanka Bhanudas Parekar, Shivraj Sangapa Shivpuje. Phytochemical Screening, Physicochemical Analysis of Starch from Colocasia Esculenta, *NeuroQuantology*, 2022; 20(20): 903-917.
78. Priti D. Mane-Kolpe, Alfa A. Jain, Tai P. Yele, Reshma B. Devkate, Priyanka B. Parekar, Komal T. Sul, Shivraj S. Shivpuje. A Systematic Review on Effects of Chloroquine as a Antiviral against Covid-19, *International Journal of Innovative Science and Research Technology*, 2022;7(11): 989-995.
79. Dr. Rohit Jadhav, Prof. Abhay D. Kale, Dr. Hitesh Vishwanath Shahare, Dr. Ramesh Ingole, Dr Shailesh Patwekar, Dr S J Wadher, Shivraj Shivpuje. Molecular Docking Studies and Synthesis of Novel 3-(3- hydroxypropyl)-(nitrophenyl)[1,3] thiazolo [4,5-d] pyrimidin2(3H)-one as potent inhibitors of P. Aeruginosa of S. Aureus, *Eur. Chem. Bull.* 2023; 12(12): 505-515.
80. Priyanka B. Parekar, Savita D. Sonwane, Vaibhav N. Dhakane, Rasika N. Tilekar, Neelam S. Bhagdewani, Sachin M. Jadhav, Shivraj S. Shivpuje, Synthesis and Biological Evaluation of Novel 1,3,4-Oxadiazole Derivatives as Antimicrobial Agents, *Journal of Cardiovascular Disease Research*, 2023; 14(8):611-624.
81. Kavita R. Mane, Prachi A. Ghadage, Aishwarya S. Shilamkar, Vaishnavi A. Pawar, Sakshi B. Taware, Priyanka B. Parekar, Shivraj S. Shivpuje. Phytochemical Screening, Extraction and In-vivo study of Immunomodulation effect of *Withania somnifera*, *Momordica dioica* and *Annonasquamosa* leaves. *Journal of Cardiovascular Disease Research*, 2023; 14(9): 231-241.